

Hypoglycemia Enhances Bupivacaine-induced Cardiotoxicity in the Rat

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The effect of blood glucose concentration on bupivacaine-induced cardiotoxicity was investigated in normoglycemic and hypoglycemic adult rats and compared to that of equipotent doses of lidocaine. The anesthetic agents were injected intraperitoneally into tracheostomized animals anesthetized with ketamine. ECG and direct blood pressure measurements were recorded continuously. Femoral arterial blood was used for determinations of glucose level, potassium concentration and base deficit values. Blood was drawn from the heart at the time of death for local anesthetic levels. In hypoglycemic animals, bupivacaine rapidly produced serious dysrhythmias leading to asystole. In normoglycemic rats, only ST-segment changes followed bupivacaine injection and death ensued from hypoxemia secondary to respiratory failure. With lidocaine, both hypoglycemic and normoglycemic rats died of hypoxemia following respiratory paralysis without antecedent dysrhythmias. Thus, hypoglycemia enhanced the cardiac effects of bupivacaine but not those of lidocaine. (Key words: cardiotoxicity: bupivacaine vs lidocaine, normoglycemia vs hypoglycemia)

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The clinical impression that the threshold for circulatory collapse consequent to intravascular injection of bupivacaine is lower in pregnant than nonpregnant patients¹ has been confirmed in sheep experiments^{2,3}. Pregnancy favors the development of "accelerated starvation" (predisposition to hypoglycemia and ketogenesis) after even short periods of fasting⁴. Dextrose containing intravenous solutions are frequently avoided for hydration prior to regional block to prevent fetal

hyperglycemia and consequent neonatal hypoglycemia. Such prehydration, however, is often administered in the absence of a current maternal blood glucose determination. Since low blood glucose levels tend to produce cardiovascular instability⁵, we suspected that hypoglycemia might be a contributing factor for the more serious cardiotoxicity noted in obstetric patients.

Rats have been used as a model for diabetes research because their glucose metabolism is comparable to that of man⁶. Rats have also been employed for studies of the effect of local anesthetics on the heart⁷. We therefore chose the rat as a model for an investigation of the influence of blood glucose

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concentration on the cardiovascular effects of bupivacaine and lidocaine.

Methods

Following approval by our Animal Investigation Committee, the cardiac effects of bupivacaine and lidocaine were assessed in normoglycemic and hypoglycemic Wistar rats anesthetized with ketamine. Preliminary studies were undertaken in normoglycemic animals to elicit the appropriate doses of the respective drugs. The lowest dose of ketamine to produce unconsciousness was found to be 80 mg·kg⁻¹ body weight (b.w.) and the lowest dose of bupivacaine to universally initiate ECG changes was observed to be 100 mg·kg⁻¹ b.w. Ketamine 80 mg·kg⁻¹ b.w. was therefore chosen as sleep dose for all rats and bupivacaine 100 mg·kg⁻¹ b.w. was selected for all experiments dealing with this agent. Lidocaine, in contrast, was used in three different doses (200, 300, 400 mg·kg⁻¹ b.w.) because of diversities in the reported potency ratios between bupivacaine and lidocaine in different species^{3,11-13}.

After establishment of anesthesia, a tracheostomy was performed in order to ensure a patent airway and to permit suctioning. The left femoral artery was cannulated for continuous blood pressure (BP) recording and intermittent blood gas determinations. Electrocardiographic (ECG) leads were applied for constant recording of cardiac activity. Blood drawn from the arterial line immediately before local anesthetic administration was used for determinations of blood glucose level, potassium concentration and base deficit value (calculated from pH and Pco₂). The local anesthetic was injected into the peritoneal cavity over three minutes using a weight-based dosage.

The investigation was carried out in two phases on adult nonpregnant female

rats of similar age (60-70 days). The first phase was conducted on sets of three animals: One rat (control) was fed until approximately one half hour prior to the study, when food was removed from the cage. The second animal was fasted for one and a half days with unrestricted access to water and a third was fed but injected with regular insulin (1 unit·kg⁻¹) one half hour before the experiment. Bupivacaine 100 mg·kg⁻¹ was used in 7 sets; lidocaine was administered in 9 sets as follows: 200 mg·kg⁻¹ in four, 300 mg·kg⁻¹ in three, and 400 mg·kg⁻¹ in two sets. At the moment of cardiac asystole, blood was drawn from the heart for determination of local anesthetic concentration in 4 of the sets of rats receiving bupivacaine and 3 of the sets receiving lidocaine 200 mg·kg⁻¹. The samples were drawn into heparinized tubes which were immediately placed on ice. The plasma was removed by centrifugation and frozen until assayed by chromatography/spectrometry for the two local anesthetics and their metabolites: 2,6-pipecolylylidine (PPX) for bupivacaine and monoethylglycinexylidene (MEGX) for lidocaine^{8,9}.

The second phase was undertaken to determine blood bupivacaine levels in normoglycemic rats corresponding to the time of death of the hypoglycemic animals. Three fed rats were sacrificed by an overdose of pentobarbital 5 min after a 100 mg·kg⁻¹ bupivacaine injection and blood was immediately obtained from their heart.

The data are expressed as the mean ± the standard error of the mean. Analysis of variance (ANOVA) and the linear coefficient of correlation were used for the statistical analyses. Differences were considered statistically significant if the calculated *P* value was less than 0.05.

Results

All rats were of similar weight (216

Table 1. Details of the ECG changes in the bupivacaine-injected rats
 Note that the interval between injection of bupivacaine and onset of ECG
 changes is expressed in seconds in the hypoglycemic animals and in minutes in
 the controls.

#	Group of rats																
	Starved					Insulin					Control						
	ECG 1	ECG 2	ASYST	ECG 1	ECG 2	ASYST	ECG 1	ECG 2	ASYST	ECG 1	ECG 2	RR ϕ	ASYST				
	Dx	sec	Dx	sec	#	Dx	sec	Dx	sec	#	Dx	min	min				
1	ST↓	185	bigem	240	330	1	HR↓	240	HR↓↓	350	1	HR↓	6	ST↓	10	13	14
2	PVC↑	210	PVC↑↑	235	305	2	HR↓	185	HR↓↓	245	2	HR↓	7	ST↓	9	13½	15
3	QRS>	175	PVC↑	245	295	3	QRS>	170	PVC↑↑	210	3	HR↓	7	HR↓↓	10	11½	13
4	HR↓	180	HR↓↓	250	315	4	ST↓	180	bigem	205	4	HR↓	6	ST↓	8	13	14
5	ST↓	240	bigem	265	345	5	HR↓	210	HR↓↓	275	5	HR↓	4½	ST↓	7½	12	13
6	PVC↑	180	PVC↑↑	230	320	6	PVC↑	230	PVC↑↑	260	6	HR↓	5	ST↓	9	13½	15
7	QRS>	175	bigem	255	350	7	bigem	240	bigem	260	7	HR↓	7	ST↓	7	12	13
8	HR↓	190	HR↓↓	240	310	8	ST↓	190	bigem	305	8	HR↓	6½	ST↓	8	11½	13
9	QRS>	235	PVC↑↑	335	390	9	PVC↑	215	PVC↑↑	265	9	HR↓	5	ST↓	9	13	14

ECG 1 = initial ECG changes
 ECG 2 = subsequent ECG changes
 Dx = diagnosis
 RR ϕ = respiratory arrest
 ASYST = asystole
 bigem = bigeminy
 HR↓ = mild bradycardia
 HR↓↓ = severe bradycardia
 ST↓ = ST segment depression
 QRS> = widening QRS complex
 PVC↑ = occasional premature ventricular contractions
 PVC↑↑ = frequent premature ventricular contractions

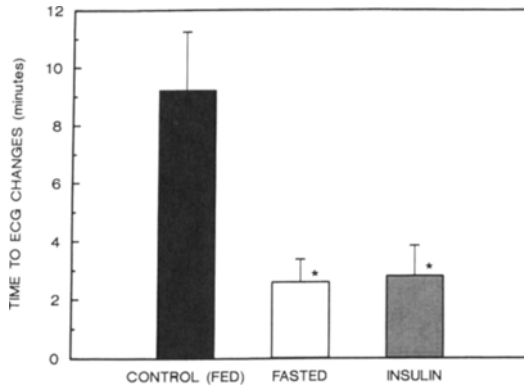


Fig. 1. Time to development of dysrhythmias consequent to bupivacaine injection.

Time (mean \pm SE) was significantly shorter for the hypoglycemic rats as compared with the control animals (* $P < 0.005$).

± 23 gm). Mean blood glucose levels in the fed animals were 9.8 ± 1.1 $\text{mmol}\cdot\text{l}^{-1}$ (normal 8.5 ± 0.9)¹⁰, in the fasted rats 4.2 ± 1.3 $\text{mmol}\cdot\text{l}^{-1}$ and in the insulin rats 4.5 ± 1.7 $\text{mmol}\cdot\text{l}^{-1}$ ($P < 0.001$ between the fed and the hypoglycemic rats). Potassium levels were within normal limits ($5.7\text{--}6.0$ $\text{mEq}\cdot\text{l}^{-1}$) in all animals. Mean base deficits were 3.7 ± 0.5 $\text{mEq}\cdot\text{l}^{-1}$ in the fed rats, 8.2 ± 0.6 $\text{mEq}\cdot\text{l}^{-1}$ in the fasted rats and 4.5 ± 0.8 $\text{mEq}\cdot\text{l}^{-1}$ in the rats who received insulin. There were no statistically significant differences between the three groups with respect to initial BP, PO_2 or PCO_2 .

In the fed animals, the injection of bupivacaine was followed by ST-segment changes after 7–10 min. Asystole secondary to progressive respiratory depression occurred in 13–15 min. In the rats made hypoglycemic by either fasting or insulin administration, bupivacaine produced hypotension, bradycardia, ST-segment changes and serious dysrhythmias such as premature ventricular contractions and bigeminy, within 3–4 min, leading to asystole in 5–7 min (table 1). The cardiac differences in time to onset of ECG changes (fig. 1) and occurrence of asystole between normoglycemic and

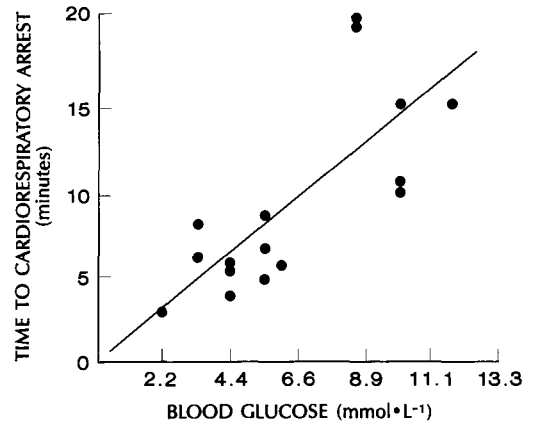


Fig. 2. Time to cardiorespiratory arrest vs. serum glucose level.

The linear coefficient of correlation (r)=0.76.

hypoglycemic animals were statistically significant ($P < 0.005$ for ECG changes and $P < 0.0001$ for death). The lower the blood glucose levels were, the shorter was the time to cardiorespiratory arrest ($r=0.76$; $P < 0.001$) (fig. 2). With lidocaine, there was no significant difference between the normoglycemic and hypoglycemic rats. All the animals in this group died of respiratory paralysis, preceded by bradycardia without dysrhythmia. With the 300 and 400 $\text{mg}\cdot\text{kg}^{-1}$ doses, slowing of the heart rate began 5–6 min after injection, with the 200 $\text{mg}\cdot\text{kg}^{-1}$ dose, 7–9 min after injection; death invariably occurred 3–5 min after the onset of bradycardia. Convulsions were not observed in any of the animals.

Blood bupivacaine concentrations ranged from 7.1 to 14.0 $\mu\text{g}\cdot\text{ml}^{-1}$ in the fed rats, from 93.3 to 216.6 $\mu\text{g}\cdot\text{ml}^{-1}$ ($P < 0.02$ vs control) in the fasted rats and from 88.8 to 153.9 $\mu\text{g}\cdot\text{ml}^{-1}$ ($P < 0.005$ vs control) in the rats receiving insulin (table 2). There was no statistically significant difference in the serum bupivacaine levels obtained between the fasted or insulin groups ($P > 0.25$). These concentrations were inversely related to the time of death ($r=-0.73$). There was a weak correlation with the plasma glucose levels

Table 2. Concentrations (mean \pm SE) of bupivacaine (4 sets of rats) and lidocaine (3 sets of rats) in cardiac blood obtained at the time of death

Group	Concentration ($\mu\text{g}\cdot\text{ml}^{-1}$)	
	Bupivacaine	Lidocaine
FED (control)	8.8 \pm 2.2	186 \pm 72
Fasted	138.4 \pm 43.1*	152 \pm 71
Insulin	96.7 \pm 26.2*	182 \pm 47

* $P < 0.05$ vs control

($r^2=0.58$).

In the three fed animals sacrificed at five minutes after the bupivacaine injection, i.e., at the usual time of death of the hypoglycemic rats, the bupivacaine concentrations were low ($10.5 \pm 2.0 \mu\text{g}\cdot\text{ml}^{-1}$). In contrast to the bupivacaine results, the lidocaine blood concentrations of all three groups were within the same range (table 2). PPX and MEGX concentrations were low in all animals. PPX ranged from 0.02 to $0.08 \mu\text{g}\cdot\text{ml}^{-1}$, MEGX ranged from $1.37\text{--}2.77 \mu\text{g}\cdot\text{ml}^{-1}$.

Discussion

Lidocaine was chosen as the comparative local anesthetic because it has been used for this purpose in many animal experiments^{2,7,13}. Furthermore, lidocaine and bupivacaine are the two most commonly employed anesthetic drugs for obstetric extradural block^{11,12}. As mentioned earlier, the reported potency ratio between bupivacaine and lidocaine has not been uniform. In humans, bupivacaine is three to four times more potent than lidocaine^{10,11}, yet in sheep and mice, the potency of bupivacaine is only twice that of lidocaine^{2,13}. Although our clinical comparisons of the two local anesthetics were undertaken at three different ratios 1:2, 1:3 and 1:4, the pharmacologic determinations were limited to the 1:2 ratio, the ratio

equipotent in mice¹³.

Our findings are comparable to those of a study evaluating the cardiac effects of intravenously administered equivalent doses of lidocaine and bupivacaine in unanesthetized sheep². After intravenous lidocaine injection, no animal developed arrhythmias other than minimal ST-T wave changes. After injection of bupivacaine, all animals had ECG changes and/or arrhythmias, most commonly widening of the QRS complex, but also premature ventricular contractions and severe ST-T wave depression.

The presence of hypoglycemia enhanced the cardiac effects of bupivacaine, regardless of whether it was produced by fasting or by insulin administration. One may thus assume that the hypoglycemic state was the primary factor causing a reduced threshold for cardiac toxicity.

Bupivacaine levels in the heart of the normoglycemic rats were low regardless of whether death had occurred by local anesthetic induced cardiac arrest (13–15 min after the bupivacaine injection) or the animals were sacrificed by administration of pentobarbital (5 min after the bupivacaine injection). In the hypoglycemic rats, by contrast, the levels were highest in the animals dying the fastest. There appear to be two possible mechanisms for the increased drug levels. One is a reduction in cardiac output with retention of unmetabolized drug, the other is an alteration of unknown nature in the metabolic degradation of the drug. Whatever the explanation, such high levels may contribute to the difficulty in cardiopulmonary resuscitation following accidental intravascular injection of bupivacaine in healthy patients. The absence of convulsions is explained by the presence of an anesthetic state and/or the use of a local anesthetic large enough to paralyze the central nervous system without excita-

tion.

Bupivacaine has been shown to cause significant changes in cardiac conduction and repolarization^{14,15}. Glucose depletion is known to attenuate normal cell repolarization¹⁶. In addition, an *in vitro* study on mechanisms of bupivacaine-induced depression of myocardial contractility suggested that inhibition of energy metabolism was a major contributor to bupivacaine cardiotoxicity¹⁷. Although only the role of calcium chloride and adenosine triphosphate was assessed, glucose is an essential provider of energy for aerobic metabolism. Thus, it appears that hypoglycemia may be added to the list of other factors, such as hypoxia, hypercarbia or acidemia, which are known to accentuate the cardiac effects of bupivacaine.

Whether the above findings can be extrapolated to the human remains questionable. However, the study has demonstrated an important difference in the cardiac effects of bupivacaine versus lidocaine.

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